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Metabolic engineering of *Picea abies* for receptor mediated induction of fluorescence and olfactory signaling

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Abstract

Reoccurring seasonal harvesting of Picea abies (Norwegian spruce) and the subsequent attachment of light fixtures to the tree followed by composting after only 3 weeks of minimal use is not only a waste but also leads to diminishing spruce forests, release of carbon dioxide, global warming, increased energy consumption and a political burden to the ongoing diplomatic efforts of Santa Claus. We here present a synthetic biology solution to address this key concerns for winter celebrations – A song induced Christmas tree with endogenous light and odor emission. Advances in synthetic biology, tissue engineering and metabolic engineering have now provided new insights and techniques for controlling whole organism signaling and bio-compound production *in situ*, and recent efforts in synthetic biology have demonstrated that complex regulatory and metabolic networks can be designed and engineered in microorganisms and simple eukaryotes. Leading researchers at the NorthPole Bioscience Engineering Council here disclose how sonic-induced signals received through tissue engineered tympanal organs are incorporated into regulating the metabolic distribution of fluorescent and olfactory signals in *Picea abies.* Upon induction, the cellular nodality of the tree branches emit light at predefined wavelength due to engineered luciferases mTwinkie (yellow), mWonderbread (white) and mSnoBalls (pink). The synthetic induction system also triggers the production and emission of seasonal olfactory signals including trans-cinnamaldehyde (cinnamon), 6-gingerol (ginger), and methyl salicylate (mint). The synthetic pathway is hereby denoted *xmas1*.

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Introduction

In the current political climate where Santa and the elves are being challenged by increasingly outspoken non-believers who propose that there is no Santa (Myers 2005) it is instrumental for the North Pole region to emphasize how Christmas can be properly integrated within the social fabric of the industrialized and developing nations through new technology development and social responsibility (Johnson and Johnson North The Pole Economic 2003). Development Council has accordingly made a concerted effort to roll out financial and regulatory support for expansion into the areas of synthetic biology, systems biology, tissue engineering, metabolic engineering and cleantech with a focus on near-term commercial applications. The North Pole has accordingly been declared a HUBZone facilitate the necessary financial to investments (Geithner, Obama et al. 2008).

There are approximately 500,000 acres of Christmas trees (usually Abies alba, Abies procera, Abies plastica and related species) planted in the US alone (Silver and Fir 2006). With a tree growth cycle of 10 years, the resources required to fulfill the seasonal need of pine scent is significant (Deere, Massey-Ferguson et al. 2008). Global costs associated with the rearing, upkeep and recycling of pine trees used for the short holiday season also include the annual release of 2,430 metric ton of CO₂ and an estimated 28% increased energy consumption to light up the trees during holiday season (Alka and Seltzer 2001).

We here present a synthetically engineered *Picea abies* (Norwegian spruce). Sonic sensors were transplanted from the tympanal organ of the African locust (*Schistocerca gregaria*) and tuned to the wavelength of children emitting shrills. The signal cascades via cinnamaldehyde-induced apoptosis in *Picea* PLC/PRF/5 cells through activation of the proapoptotic Bcl-2 family proteins and MAPK pathway and activate a set of luciferases emitting light at three different wavelengths. The luciferases are clustered at the node of each tree branch. The same tympanal organ signal transduction pathway also activates the release of trans-cinnamaldehyde (smell of cinnamon), 6-gingerol (smell of ginger) and methyl salicylate (mint-like smell).

Materials ands methods

Synthetic genes were designed by and purchased from DNA2.0 (Menlo Park, CA). Growth media and standard molecular biology buffers were purchased from Teknova (Morgan Hill, CA). Restriction enzymes were purchased from New England Biolabs (Ipswish, MA). All materials and chemicals not otherwise noted were purchased from The Elves Workshop Supplies Inc. (Santa's Secret Village, NorthPole) and were of analytical grade or equivalent.

Heterologous expression host

Picea abies strain Marve (F^- mcrA Δ (mcrBC-hsdRMS-mrr) recA1 endA1 lon gyrA96 thi supE44 relA1 $\lambda^{-} \Delta(lac-proAB)$ (Fleksnes and Wesenlund 1972) was used as host for the engineering of the synthetically introduced *xmas1* pathway. The Norwegian spruce was chosen for having a metabolic network that could easily be deceived into accepting non-natural biological building blocks. Truth be told, the Norwegian spruce is clearly the stupidest of spruces (Keillor 2005). This unique Norwegian characteristic is very helpful when re-engineering the underlying framework without disclosing any information to the host system (see also (Opsahl 1974)).

Molecular biological procedures

Standard procedures were used for cloning, manipulation and analysis of DNA, PCR, electroporation, and transformation

Synthetic biology engineering of Christmas tree. S. Claes *et al.*,

(Sambrook, Fritsch *et al.* 1989). All oligonucleotide synthesis and DNA sequencing was performed by DNA2.0 using instruments and consumables from Applied Biosystems (Foster City, CA).

Sequence analysis

DNA sequence tracefiles were analyzed, automatically assembled into full length contiguous sequences and finished using the Staden package (Staden, Judge et al. 2001). Gene DNA sequence design and codon optimization was carried out using the Gene Designer software (Villalobos, Ness et 2006). Homology searches al. were Blast algorithm performed with the (Altschul, Madden et al. 1997) implemented on a local server.

Induction Assay

Sonic induction of the synthetic system was carried out using 109 children ranging from 4 to 8 years of age (*Cantatrix falsetto*) singing various Christmas carols (and 'This land is your land'). Extended noise level was sustained by dispersing Sour Patch Kids (Cadbury Adams, Whippany, NJ) and Tootsie Rolls (Tootsie Roll Industries, Chicago, IL). The children were furnished by the Fletcher-Maynard Elementary School (Cambridge, MA). Anecdotal evidence suggested these children to be able to emit extraordinarily high-pitched sounds (H.H. data not shown). The researcher utilized noise-canceling headphones (Panasonic RP HC500) and halothane vapor induced anesthesia during the induction assay. No child was harmed during the experiment.

Results and discussion

Sonic reception system

Signal input to the synthetic *Picea abies* system was achieved through orthogonal transplantation of insect tympanal ostium tissue and the immediately adjacent tracheal tubing to the tapetal layer

in the innermost of four sporophytic layers encapsulating the meristem of the plant. Tympanal organs are the hearing organ in insects, consisting of a membrane stretched across a frame backed by an air sac. Sounds vibrate the membrane, and the vibrations are sensed by the chordotonal organ. We reasoned that the tympanal and chordotonal organs from the infamous African locust should Schistocerca gregaria be а conductive inducer of stimuli considering the breadth, history and scope of the swarming locust (Mohammed 1787; Joel 1924). It is clear that the communication signal system of the locust must be well developed so that it can both coordinate the swarming process and sing 'When You Wish upon a Star', although not necessarily simultaneously (Cricket and Disney 1940).

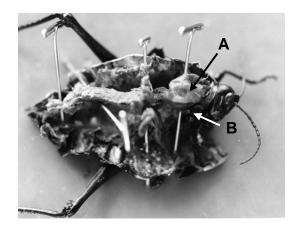


Fig.1. Dissected *Schistocerca gregaria*. The tympanal organ (A) and the underlying chordotonal organ (B) that transmit the signal is clearly visible in the anterior of the insect. Glenfiddich anesthesia (Coca and Cola 2001) was utilized during the dissection process to constrain the movement of the tissue donor

The tympanal and chordotonal tissue from 96 geographically separated locust individuals were HLA typed with predominant type I pneumocytes on frozen sections. Locust donors with alveolar epithelium expressed HEA-125, class I MHC. antigens HLA-DR (total of four individuals) were chosen as donors of tympanal and chordotonal tissue. The assumption was that this would mitigate any cerebral functions in the host. The dissected donor (Fig 1) did not object. Organs were subsequently transplanted into the meristem of *Picea abies* using previously described procedures (Einstein, Zweistein *et al.* 1974). The tree was treated aggressively with immunosuppressive agents including Cyclosporin, Tacrolimus (FK506), and Rapamycin to minimize the rejection of the tympanal and chordotonal tissue transplants.

Emitting seasonal light

The luciferase gene used in this study was originally derived from a small population of reindeer (*Rangifer tarandus*) having red noses (Pret & Zel 1995; Stroganoff and Lindström 2004). The light emission spectra of the wild type gene was engineered using the ProteinGPS technology of DNA2.0 to display light in the yellow (mTwinkie), pink (mSnoBalls) and white (mWonderbread) spectra as previously described (Hostess, Tour *et al.* 2003; Liao, Warmuth *et al.* 2007).

The three monomeric luciferase analogs where codon optimized for maximal protein expression level in Picea abies chloroplasts using the Welch algorithm (Gustafsson, Govindarajan et al. 2004). The synthetic constructs where preceded by a chordontal inducible CMV promoter and a Shine-Dalgarno (SD) sequence (GGAGG) located 4 to 12 nucleotides upstream of the initiator AUG. The monomeric luciferase genes were immediately followed by another SD and genes encoding the leaf Npr1 signaling pathway (Gross and Simmons 2005). This construction ensure that the dicistronic operon is to be expressed only at the node of each branch as it emits the light. The vector also encoded kasugamycin selection resistance for purposes.

Plastid transformation offers several unique advantages compared with nuclear genome transformation, such as high level of transgene expression within plastids, expressing multiple transgenes as operons, lack of position effect due to site-specific transgene integration, and reducing risks of gene flow via pollen due to maternal inheritance of the plastid genome.

Chloroplast containing *Picea abies* tissue was transformed with the vectors expressing the monomeric luciferase constructs using standard agrobacterium transfection methodology.

Emitting seasonal odor

The formation of cinnamaldehyde and gingerol are related to the phenylpropanoid metabolic pathways, *i.e.* the transformation of phenylalanine to cinnamic acid and its hydroxylated derivatives coumaric acid, coffeic acid, gingerol and ferulic acid. We used a phenylalanine oxidase (CinA) and a phenylalanine deaminase (CinB) identified in the wasp genome (Nasonia vitripennis) to oxidize the carboxylic group to an aldehyde to remove the amine from and phenylalanine. The GenBank sequence NM 001129280 identifier and XM 001601513 denotes the original open reading frames. The amino acid sequences were codon optimized for maximal protein expression level in *Picea abies* chloroplasts using the Welch algorithm (Gustafsson, Govindarajan et al. 2004).



Fig.2. *Picea abies* strain Marve encoding the expressing mTwinkie, mSnoBalls and mWonderbread monomeric luciferase genes. The tree was exposed to 30 minutes of children singing Christmas carols. Maximal induction could be seen after 1hr.

The enzymes CinA and CinB where engineered for increased Kcat/ K_M and

substrate specificity so that ~30% of all nonbound phenylalanine in the cell would be converted to trans-cinnamaldehyde and released through the stoma in conjunction with transpiration.

The gingerol synthesis was driven by GinT, and enzyme originating from the ginger plant (*Zingiber officinal*) where the coumarate is reduced and hydroxylated to form Gingerol (the ginger smelling compound). GinT and its enzymatic properties have previously been described (Thomson, Preston *et al.* 2007). Similarly to the CinA and CinB enzymes, the GinT enzyme was engineered and expressed at sufficient levels through codon optimization.

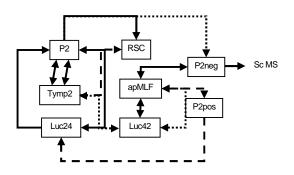


Fig.3. Tentative model describing the signal transduction pathway for the *xmas1* control system. Solid lines signifies activation, dotted lines signifies inhibition, dashed lines signifies interrogatory functionality. The model is further described in the text and elsewhere (Wait and See 2008).

Conclusion

Synthetic biologists engineer complex artificial biological systems to investigate natural biological phenomena and for a variety of applications. We here outline an immediate application of synthetic biology that integrates many levels of control and signal transudation with the goal of reducing the environmental impact of Christmas.

We specifically show methods for successfully designing and constructing

engineered cells with novel functions in a framework of an abstract hierarchy of biological devices, modules, cells, and multicellular systems. То achieve predictability and reliability, strategies for engineering Picea abies must include the notion of cellular context in the functional definition of devices and modules, use rational redesign and directed evolution for optimization, and focus system on accomplishing tasks using cell populations rather than individual cells.

Christmas trees with endogenous emission of light and smell that is induced by the sound of singing children projects the future for ongoing efforts at Santa's laboratories[©]. Future efforts include the incorporation of sound emission from the tree. This goal has so far been proven elusive as our intent to clone the famous Ho Ho Ho from Santa himself has failed despite several serious attempts. We now have lowered our initial goal and instead are proposing the cloning and heterologous expression of the sound derived from chattering elves. We are currently exploring different methods for grinding up the elves to isolate the relevant factors and expect to be able to report this exciting extension of our synthetic biology Christmas tree in due course.

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